# oroboros instruments high-resolution respirometry

Course on High-Resolution Respirometry

IOC44. Mitochondrial Physiology Network 12.24: 1-8 (2007)

# 44<sup>th</sup> International Course on High-Resolution Respirometry



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# 12-16 Dec 2007

The 44<sup>th</sup> O2k-Course on High-Resolution Respirometry includes experiments with permeabilized muscle fibers (horse skeletal muscle), providing a practical overview of the **Oxygraph-2k**, with integrated on-line analysis by **DatLab 4.2 (new update)**, and



*Right*: Dr. Dominique Votion (Belgium), veterenary specialist on horse muscle pathophysiology, at an O2k-experiment (IOC36).



demonstration of the **TIP-2k**. Emphasis will be placed on hands-on applications by all participants and a general introduction into O2k-high-resolution respirometry.

Schröcken, Vorarlberg, Austria

Experienced tutors guide small working groups step-by-step through the approach

of high-resolution respirometry. Five Oxygraph-2k, three TIP-2k and several PCs are available for a do-it-yourself application of both hardware and software.

During lunch breaks, sufficient time is available for relaxing walks and talks, skiing and snow shoe walking, to enjoy the refreshing

scenery of the alpine environment, or use the spare time for specific tutorials.

Snowfall may contribute to or interfere with outdoor activities, but performance of the OROBOROS Oxygraph-2k is weatherindependent. With DatLab 4.2 we accomplish data analysis on-line during the experiment, providing final results and their graphical presentation by the end of an experimental run. Thus we gain sufficient time to see the Titration-Injection microPump TIP-2k with new feedback-control in action and practice its simple and automatic operation.



www.oroboros.at

# **Support MITOFOOD** COST Action Number FA0602 (Coordinator: Dr. Jaap Keijer, RIKILT-Institute of Food Safety, Wageningen University, The Netherlands.

## Tutors

Med. Univ. Innsbruck, Dept. General Transplant Surgery, D. Swarovski Research Lab., Innsbruck, Austria; and OROBOROS INSTRUMENTS, Austria

- **Fasching Mario**, PhD, mario.fasching@oroboros.at (*tutor*)
- **Gnaiger Erich**, PhD, erich.gnaiger@i-med.ac.at (*lecturer* & *tutor*)
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- **Lemieux Hélène**, PhD, helene.lemieux@oroboros.at (*lecturer* & *tutor*)
- **Scandurra Francesca**, PhD, francesca.scandurra@oroboros.at (*lecturer & tutor*)
- **Subarsky Patrick**, PhD, patrick.subarsky@oroboros.at (*tutor*)

# **Guest lecturer**

• Anderson Ethan J., PhD, Department of Exercise and Sport Science & Department of Physiology, Brody School of Medicine, East Carolina University, Greenville, NC, USA.

## Programme

## Wednesday, 12. December

Participants arriving in Bregenz: Meeting					
		point at Bregenz train			
		station,	1	L.1	hour
		drive to Schröcken.			
Aftern	oon/Evening	Check	in	at	Hotel
		Mohnen	fluh		
19:30	Dinner				
		After dinner, Walcomer			



After dinner: Welcome; introduction of participants.

### Day 2: Thursday, 13. December

- 08:45 11:45 Principles of high-resolution respirometry from switching on the Oxygraph-2k to the experimental result.
  - Oxygraph-2k demo experiment with DatLab 4.2 (new update).
  - Oxygen calibration of the polarographic oxygen sensors (POS).



• Preparation of permeabilized muscle fibers.

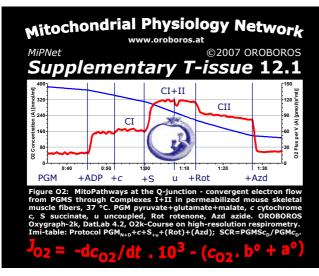
 Determination of fiber wet weight with <u>METTLER TOLEDO</u> microbalance <u>XS205DU</u> (1 to 3 mg per chamber).



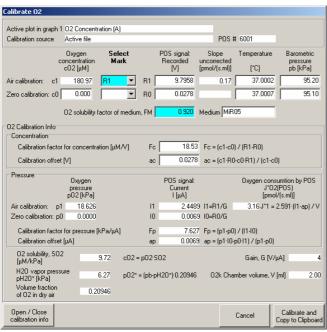
- Addition of permeabilized skeletal muscle fibers, closing the chamber.
- Multiple substrate OXPHOS titration protocol: ADP control ratio, succinate control ratio, phosphorylation control ratio.

#### 12:00 - 16:00

Lunch break; skiing (bus leaves at 12:22 from Hotel Mohnenfluh).



# Day 3: Friday, 14. December



16:15 -19:00 Working group session 1: Hands-on with the Oxygraph-2k (four instruments - eight parallel chambers): Preparation of permeabilized skeletal muscle fibers; OXPHOS titration protocols. On-line DatLab analysis. Q 19:30 Dinner 21:00 MiPNet Session 1. Erich Gnaiger (Innsbruck, AT): Highresolution respirometry and OXPHOS titration protocols: Flux Control Ratios. **Discussion of results.** 

08:45 - 11:45 Working group session 2: Hands-on experiments with the

Oxygraph-2k - instrumental performance: O2k-calibration and background test, on-line DatLab analysis.

12:00 – 16:00 Lunch break, skiing

**16:15 - 19:00 Working group session 3:** High-resolution respirometry and

DatLab 4; Instrumental setup and service.

Parallel special interest group: DatLab 4 Analysis and respiratory protocols.

19:30 Dinner

21:00

Hot topics in Mitochondrial Physiology: MiPNet Session 2. Ethan J. Anderson (Greenville, US): Excess dietary fat depletes glutathione, increases mitochondrial oxidant emitting potential and shifts redox balance to a more oxidized state in human skeletal muscle.

21:30

**Hélène Lemieux** (Innsbruck, AT) Respirometry with permeabilized cultured cells: Method and application.

## Day 4 (Saturday, 15. December)



Working group session 4: Highresolution respirometry and DatLab 4; Instrumental setup, POS service.
Parallel special interest group: TIP-2k titration and injection, feedback control and steady-state.



- Special Topic: Francesca Scandurra (Innsbruck, AT) Oxygen limitation of cellular respiration: Aerobic-anoxic transitions and hypoxic steadystates.
- 13:00 17:00 Snowshoe walk to a welcome at the Alpmuseum uf m Tannberg (*we keep the details of timing flexible according to weather conditions*).



Alpmuseum uf m Tannberg, Batzen www.alpmuseum.at



### 17:30 -19:30 Working group session 5

Open topics: Problems and solutions. Special interest groups: Inhibitor titration and flux control / Experimental regimes / Oxygen kinetics.

20:00 Dinner

22:00 Discussion - Summary – Conclusions

Sunday, 16. December



Departure to Bregenz and Innsbruck

Tuesday, 18. December - OROBOROS MiPart - Dec. 18, 18:18, Schöpfstr. 18

### **CONTENTS: OVERVIEW ON HIGH-RESOLUTION RESPIROMETRY**

# Introduction: Mitochondrial and cellular respiratory physiology – new challenges for high instrumental performance.

# High-resolution respirometry – what makes the difference? Presentation of the OROBOROS Oxygraph-2k

- Low oxygen and measurement of cellular oxygen consumption pushing the limits of detection.
- Optimum system design the OROBOROS Oxygraph-2k.
- DatLab 4.2: on-line recording of oxygen concentration and flux; linear slope versus oxygen flux as a function of time.
- DatLab 4.2: the specialized software for high-resolution respirometry; high-resolution calibrations.

#### **OROBOROS Oxygraph-2k and TIP-2k: On-line instrumental performance**

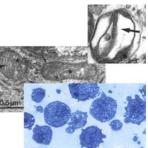
- Instrumental background: measurement and correction as a function of pO<sub>2</sub>.
  - High resolution of respiratory flux at various steady-states.
  - The Titration-Injection microPump TIP-2k: automatic titrations.
- Conceptual and methodological advantages of measurement at physiological low levels of oxygen.
- High time resolution for kinetic analyses: Determination of the time constant, dynamic corrections.

#### Polarographic oxygen sensor (POS) and O2k service

- Cleaning of anode and cathode.
- Electrolyte and membrane application.
- Oxygraph-2k and TIP-2k: instrumental maintenance.

# **Protocols for the O2k Demo Experiment**

Gnaiger E, ed (2007) *Mitochondrial Pathways and Respiratory Control.* OROBOROS MiPNet Publications, Innsbruck: 96 pp. Electronic 1<sup>st</sup> ed ISBN 978-3-9502399-0-4 – www.oroboros.at



# **Accomodation and Location**

**Hotel Mohnenfluh** www.mohnenfluh.at; Tel.: +43 5519 203; hotel@mohnenfluh.at. The course takes place at Hotel Mohnenfluh (Sylvia Schramm-Strolz, *right*), including accomodation for all participants breakfast, meals and coffee breaks.

#### Skiing



### Warth-Schröcken - http://www.snowworld.at/.

The skiing area Salober is reached by a free bus service, leaving at 12:20/12:22 at Hotel Tannberg / Hotel Mohnenfluh. The following information relates to prices in 2006/07, which may have changed: For the afternoon after 12:30, the skiing pass is  $\in$  22.50 for the skiing lifts of Salober and Warth. There is also excellent crosscountry skiing around lakes Kalbelesee and Körbersee, as well as easy walking in magnificent winter scenery. Ski rental is available in Schröcken and at the skiing lift Salober. Top ski (+boots) is  $\in$  16.- (+7.-; 1 day), 30.- (+12.-; 2 days), 42.- (+17.-; 3 days) or 52.- (+22.-; 4 days). You can return to Schröcken on skies (depending on snow conditions) or by the free bus (leaving 15:30 at Salober).

WeatherSnowfall and sub-freezing temperatures are expected in<br/>December. Sunshine may be strong – bring sunglasses and<br/>sunscreen, even if you do not plan to go skiing. Protect yourself<br/>against wind and potential snowfall or rain (gloves, jacket, etc.).

**Further information** Introductory course material is available on our homepage <u>www.oroboros.at</u>.

Oxygraph-2k

# Contact

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## **Participants**

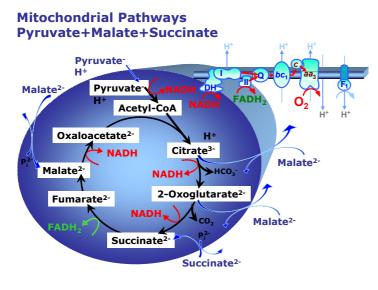
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# Hot topics in Mitochondrial Physiology –

# **MiPNet Abstracts**

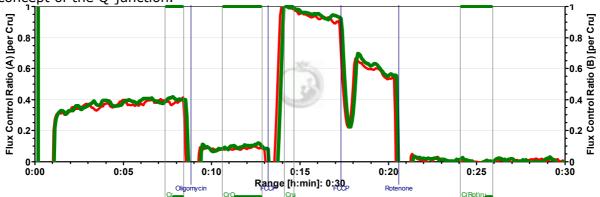


MiPNet 1. High-resolution respirometry and OXPHOS titration protocols: Flux Control Ratios.

Erich Gnaiger<sup>1,2</sup> <sup>1</sup>Medical Univ. Innsbruck, Dept. Transplant Surgery, D. Swarovski Research Lab., Innrain 66, 6020 Innsbruck, Austria. <sup>2</sup>OROBOROS INSTRUMENTS, Schöpfstr. 18, Innsbruck, Austria. erich.gnaiger@i-med.ac.at

Electron flow in the mitochondrial respiratory chain drives proton translocation through the inner mitochondrial membrane, buildina а membrane potential and proton motive force which in turn fosters the power for oxidative phosphorylation. Metabolic maps in bioenergetics carefully point out that, in contrast to a

linear arrangement of respiratory complexes, input into the electron transport chain converges from Complexes I and II (CI+II, and other flavoproteins) into the Q-cycle. The implications of this Q-junction on mitochondrial respiratory control are not sufficiently recognized in bioenergetics and metabolic flux control analysis. The Q-junction emerges now as a novel paradigm of respiratory control in mitochondrial physiology, based on high-resolution respirometry (OROBOROS Oxygraph-2k [1]) in permeabilized cells and tissue preparations. ADP-activated respiration with malate+glutamate or pyruvate (classical State 3) increases up to 2-fold after addition of succinate [1]. Parallel electron input converging at the O-junction shares flux control with the phosphorylation system, and corresponds to mitochondrial substrate supply in vivo. By establishing the reference state of maximum coupled respiration, convergent electron input into the Q-junction provides the proper basis for (i) quantifying excess capacities, metabolic thresholds, and interpreting flux control by various enzymes (e.g. COX) and functional units (phosphorylation system), and (ii) evaluation of specific enzymatic defects in mitochondrial respiratory physiology and pathology. The design is discussed of a reference protocol for multisubstrate/inhibitor titrations, which takes into account the concept of the Q-junction.



The new version of DatLab 4.2 includes an option for plotting Flux Control Ratios (FCR). Interpretation of such flux control ratios is discussed in comparison to the classical respiratory control (RCR) or uncoupling control ratios (UCR).

1. Gnaiger E, ed (2007) *Mitochondrial Pathways and Respiratory Control.* OROBOROS MiPNet Publications, Innsbruck: 96 pp.

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# <u>MiPNet 2.</u> Excess dietary fat depletes glutathione, increases mitochondrial oxidant emitting potential and shifts redox balance to a more oxidized state in human skeletal muscle system

Ethan J. Anderson, Kristen E. Boyle, Joseph A. Houmard, P. Darrell Neufer

*Exercise Science & Physiology, East Carolina University, Greenville, NC.-andersonet@ecu.edu* 

The worldwide epidemic of Type 2 Diabetes is linked to modern lifestyle, of which high dietary fat intake is a substantial component. To examine the molecular impact of a high fat diet on human skeletal muscle mitochondria, male (18-34 yrs) subjects were recruited and muscle biopsies were obtained following a 12-hour fast (A), 4 hours after a single high-fat meal (65-70 % fat content, B), and after 5 days of high-fat diet/12-hour fast (C). Using a method to measure in situ mitochondrial respiration and H<sub>2</sub>O<sub>2</sub> emission (mH<sub>2</sub>O<sub>2</sub>, an index of reactive oxygen species) in permeabilized muscle fibers supported by glutamate+malate (GM), succinate (S), and palmitoyl-carnitine (Pc), we observed from A to both B and C a striking 2.5- and 3-fold (P<0.05) increase in Vmax of G+M/S and Pc-supported mH<sub>2</sub>O<sub>2</sub> emission, respectively. Surprisingly, from A to C the total glutathione content (GSH<sub>t</sub>) was decreased by 40 % (P<0.05), and the GSH/GSSG ratio decreased by 2.5-fold (P<0.05). Respiration with all substrates remained unchanged from A to B, but basal and maximal G+M/S-supported respiration was decreased by 30 % (P<0.05) at C.

These data demonstrate that an excess dietary fat intake (1) increases skeletal muscle  $mH_2O_2$  emission and shifts redox balance to a more oxidized state, (2) decreases capacity for scavenging  $mH_2O_2$  by depleting  $GSH_t$ , (3) decreases mitochondrial respiration, all potentially contributing to the etiology of Type 2 diabetes.

