# **Oroboros** O2k-Protocols mt-Preparations

Mitochondrial Physiology Network 20.15(02):1-2 (2016)

Version 02: 2016-03-25 ©2015-2016 Oroboros

Updates: http://wiki.oroboros.at/index.php/MiPNet20.15 IsolationRatHeart-mt



# Laboratory protocol: isolation of rat heart mitochondria

Sumbalova Z<sup>1</sup>, Fontana-Ayoub M<sup>2</sup>, Krumschnabel G<sup>2</sup>

<sup>1</sup>Pharmacobiochem Lab, Fac Medicine, Comenius Univ, Bratislava, Slovak Republic

#### <sup>2</sup>Oroboros Instruments

Schöpfstr 18, A-6020 Innsbruck, Austria

Email: instruments@oroboros.at

www.oroboros.at

# 1. Preparation

Switch on centrifuge and let it cool down to 4 °C. Keep all buffers, dissection gear, homogenization tools and centrifuge rotor at 4 °C (or on ice).

#### 1.1. Anesthesia

Rats are anesthetised by intraperitoneal injection of Thiopental (0.1 g/kg) or by  $CO_2$  narcosis.

# 1.2. Isolation procedure

- 1. Kill rat, dissect out heart (take weight) and put it into ice-cold isolation medium A, wash to remove blood, discard all medium.
- 2. Cut the heart into small pieces (should become a mash), add drops of isolation buffer A while cutting.
- 3. Add isolation buffer B (10 ml/g tissue), transfer to pre-cooled glass/Teflon potter and homogenize with 8-12 strokes at medium speed (1,000 rpm).
- 4. Transfer the homogenate to a 50 ml beaker, bring volume up to  $\sim 15$  ml/g tissue with isolation buffer A, place it on a magnetic stirrer, stirr slowly for 20 min in an ice bath.
- 5. Re-homogenize the homogenate briefly in the potter, bring the volume up to  $\sim 20-30$  ml with isolation buffer A.
- 6. Centrifuge: 1,000 *q*, 10 min, 4 °C.
- 7. Transfer the supernatant to a new 50 ml Falcon tube.
- 8. Centrifuge: 6,200 *g*, 10 min, 4 °C.
- 9. Discard the supernatant, carefully re-suspend the mitochondrial pellet in a small volume of isolation buffer C and fill up to a volume of 20-30 ml (for 1 q tissue) with isolation buffer C.
- 10. Centrifuge: 6,200 q, 10 min, 4 °C.
- 11. Discard supernatant and carefully re-suspend mitochondria with small volume of isolation buffer C. The volume of mitochondrial suspension for 1 g tissue is  $\sim 1.2$  ml.
- 12. Store mitochondria on ice; perform functional studies within 3-4 h.

- 13. Transfer subsamples (20  $\mu$ l) into Eppendorf tubes and store at  $-20^{\circ}$ C for further analysis (protein concentration, citrate synthase activity).
- 14. For respiratory measurements add 2.5 μl of mitochondrial suspension into the 2 ml O2k-Chamber.

## 2. Media

## 2.1. Isolation buffer A

-		
Chemical	Final conc.	Required for
		500 ml buffer
KCI	180 mM	6.71 g
EDTA	4 mM	0.745 g
BSA	1 g/l	0.5 g

Adjust pH to 7.4 with Tris, HCl

#### 2.2. Isolation buffer B

Isolation buffer A with 0.25 mg/ml Subtilisin. Add 2.5 mg Subtilisin to 10 ml of Buffer A.

#### 2.3. Isolation buffer C

Isolation buffer A without BSA.

# 2.4. Preparation of buffers

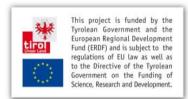
Prepare 500 ml of buffer C for 3 - 4 isolations. Add BSA to 250 ml of buffer C to obtain buffer A. A and C can be stored at -20 °C. Prepare buffer B fresh.

#### **Acknowledgements**

Sumbalova Z was supported by Action Austria -



Slovakia (2015). O2k-Protocol development supported in part by K-Regio project MitoFit. www.mitofit.org





http://wiki.oroboros.at/index.php/O2k-mitochondrial preparations