Experimental Communication

Cite

Heimler SR, Phang HJ, Bergstrom J, Mahapatra G, Dozier S, Gnaiger E, Molina AJA (2022). Platelet bioenergetics are associated with resting metabolic rate and exercise capacity in older women. MitoFit Preprints 2022.7. https://doi.org/10.26124/mito fit:2022-0007

Author contributions

GM and SD conducted data collection. EG provided technical expertise and assistance. JB, HJP, and SRH performed data analysis. SRH and HJP wrote and edited the manuscript. AJAM conceptualized the study and edited the manuscript.

Conflicts of interest

The authors declare they have no conflict of interest.

Received 2022-04-12 **Accepted** 2022-04-12 **Published** 2022-04-12

Data availability

Data available Open Access https://doi.org/10.5281/zenodo.64520

Keywords

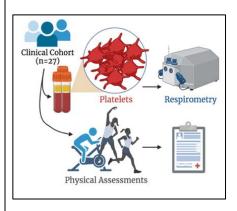
platelets; resting metabolic rate (RMR); cardiopulmonary exercise testing (CPET); OXPHOS capacity *P*; electron transfer capacity *E*; inverted regression analysis

Platelet bioenergetics are associated with resting metabolic rate and exercise capacity in older adult women

Stephanie R Heimler^{1*},
Howard J Phang^{1*},
Jaclyn Bergstrom¹,
Gargi Mahapatra²,
Stephen Dozier¹,
Erich Gnaiger³,
Anthony J A Molina^{1**}

- 1 Division of Geriatrics, Gerontology, and Palliative Care; Department of Medicine, University of California San Diego School of Medicine, La Jolla, CA, USA.
- 2 Section on Gerontology and Geriatrics, Sticht Center for Healthy Aging and Alzheimer's Prevention, Department of Internal Medicine, Wake Forest Baptist Medical Center, Winston-Salem, NC, USA.
- 3 Oroboros Instruments, Innsbruck, Austria.
- * Co-first authors
- ** Corresponding author: ajmolina@health.ucsd.edu

Abstract



This study investigates relationships between platelet mitochondrial bioenergetics and resting metabolic rate (RMR), body composition, and exercise fitness in women over 60 years of age. We report positive correlations

between peak respiratory exchange ratio (RER) and RMR with five measures of platelet respiration, supporting the premise that blood cells can be utilized to report on mitochondrial function associated with physical health and fitness. Identifying mechanisms associated with physical performance among older adults supports the development of reliable biomarkers of healthy aging and can advance the development of efficacious interventions.



1. Introduction

1.1. Age-related bioenergetic decline and physical function

Mitochondrial dysfunction is a biological hallmark of aging implicated in multiple age-related diseases and disorders, including physical function decline and sarcopenia (López-Otín et al 2013; Lenaz et al 2000; Gonzalez-Freire et al 2018). Age-related skeletal muscle bioenergetic decline is marked by decreased mitochondrial density, ATP production, electron transfer (ET) capacity, and tricarboxylic acid cycle enzyme activity (Short et al 2005; Marzetti et al 2013). This decline in skeletal muscle bioenergetics is associated with decreased cardiopulmonary fitness and exercise fitness (Coen et al 2013). For example, multiple studies found that gait speed, a clinical measure of functional capacity integrating multiple systems including the nervous system and musculoskeletal system, is associated with skeletal muscle mitochondrial bioenergetics (Choi et al 2016; Tyrrell et al 2015).

1.2. Resting metabolic rate

Although physical activity comprises 15-30 % of daily energy expenditure, the major contributor to total energy expenditure is resting metabolic rate (RMR), which accounts for roughly 60 % of the body's total energy demands (Ravussin, Bogardus 1989). RMR is largely determined by metabolically active tissues including skeletal muscle, heart, brain, kidney, liver, and — in certain populations — adipose tissue (Wang 2010). Although RMR has been extensively examined in relation with age, sex, body composition, and physical activity, few studies have examined the relationship between mitochondrial function and RMR (Larsen et al 2011; McMurray et al 2014). However, RMR was recently found to be associated with in vivo skeletal muscle oxidative capacity, suggesting a strong link between mitochondrial function and RMR (Edwards et al 2013; Zampino et al 2020).

1.3. Blood-based bioenergetic profiling

There is mounting evidence that blood-based bioenergetic profiling can be utilized to report on systemic bioenergetic capacity, and is related to mitochondrial function measured in other tissues (Nguyen et al 2019; Mahapatra et al 2018; Sjövall et al 2014). Our group has shown that blood cell respirometry correlates with skeletal and cardiac muscle respirometry (Tyrrell et al 2016). These data suggest that respirometry of blood cells may reflect the bioenergetic status of other organ systems, including skeletal muscle. The goal of this study is to uncover whether platelet respirometry correlates with RMR, body composition, and measures of exercise fitness. In particular, platelet mitochondrial function has been reported to be correlated with skeletal muscle mitochondrial function and exhibit bioenergetic changes associated with age in humans (Braganza et al 2019).

1.4. Study goals

The study presented here examines the relationship of platelet bioenergetics with resting metabolic rate and exercise capacity in community-dwelling women over 60 years of age. While women have a longer life expectancy than men, they exhibit higher rates of frailty later in life. Moreover, men continue to exhibit higher physical performance, even later in life (Hägg, Jylhävä 2021).



2. Materials and methods

2.1. Participants

Twenty-seven healthy adult women (mean age = 70.2 years) were included in this study. All participants were free from chronic medical illness, current health complaints, and abnormal physical examination (including blood pressure ≥140/90 mmHg). Screening tests, including electrocardiogram, exercise echocardiogram, and spirometry, indicated no presence of heart disease. Participants who regularly undertook vigorous exercise were excluded from this study. The protocol for this study was approved by the Wake Forest School of Medicine institutional review board, and all participants provided written, informed consent.

2.2. Resting metabolic rate

RMR was measured using indirect calorimetry (MGC Diagnostics) for each participant after an overnight fast as previously described (Nicklas et al 2019).

2.3. Exercise fitness

Exercise fitness was measured by cardiopulmonary exercise testing (CPET), an integrative assessment of exercise responses particularly useful in the context of cardiovascular or pulmonary disease (Balady et al 2010). It integrates evaluation of the pulmonary, cardiovascular, haematopoietic, neuropsychological, and skeletal muscle systems, which are not adequately reflected through the measurement of individual organ system function (Albouaini et al 2007). Ventilatory and gas exchange responses were measured on a breath-by-breath basis (MGC Diagnostics, St. Paul, MN) using a treadmill ramp protocol to exhaustion as previously described (Nicklas et al 2019).

2.4. Body composition

We recorded several body composition measurements, including: BMI, lean mass, and fat mass. We recorded lean mass and fat mass values using total body dual-energy x-ray absorptiometry (DXA) on the Prodigy Scanner (General Electric, Madison, WI) as previously described (Nicklas et al 2019).

2.5. High-resolution respirometry

Platelet mitochiondrial function was assessed by high-resolution respirometry using the Oxygraph-2k (Oroboros Instruments, Innsbruck, Austria), which provided a detailed analysis of respiratory pathway control and coupling control (Gnaiger 2020). Our

protocol is depicted in Figure 1 and was comprised of the following injections: catalase (Ctl; 112,000 U/mL), ADP (D; 0.5 mM), magnesium (Mg; 0.3 mM), digitonin (Dig; 10 mg/mL), octanoylcarnitine (Oct; 0.1 mM), malate (M; 0.1 mM and 0.8 mM), cytochrome *c* (c; 4 mM), pyruvate (P; 2 mM),

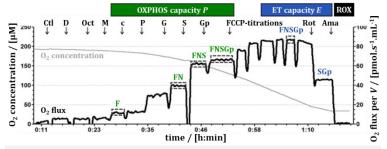


Figure 1. Representative trace of high-resolution respirometry of platelets.



glutamate (G; 2 mM), succinate (S; 1 mM), and glycerophosphate (Gp; 1 mM). After, we titrated the uncoupler FCCP in 1 μ M steps until maximal respiration is reached. Finally, we added rotenone (Rot; 1 μ M) and antimycin-A (Ama; 2.5 μ M) to stop mitochondrial respiration (residual oxygen consumption, ROX). The platelet concentration was $2\cdot10^8$ cells per 2 mL chamber.

2.6. Statistical analysis

We determined Pearson correlation coefficients r and partial correlations between respirometry measures and CPET, body composition, and calorimetry measures, including adjustments for age, BMI, and fat %. Regression lines were calculated according to inverted regression analysis (Gnaiger 2021).

3. Results

3.1. Participant characteristics and platelet respiration

This study included 27 healthy older adult women (mean age 70.2 ± 1.1 years). Platelet OXPHOS capacities *P* [pmol·s⁻¹·mL⁻¹] were for fatty acid oxidation (F_P; 19.4 ± 1.7), F-& NADH(CI)-linked P (FNP; 24.8 \pm 2.0), FN- & succinate-linked P (FNS_P; 37.6 ± 3.0), FNS- & glycerophosphate-linked P (FNSGp_P; 45.0 \pm 3.6) and the corresponding ET-capacity E(FNSGp_E; 76.6 ± 6.6 ; Table 1: O₂ flow per platelet). These respiratory states have previously been described as FAO. FAO+Complex I, FAO+Complex I+Complex II,

Table 1. Participant characteristics (N=27)

		Mean (SE)				
Demographics						
age [years]		70.2 (1.1)				
Platelet respirometry						
F_P	[amol·s ⁻¹ ·x ⁻¹]	0.194 (0.017)				
FN_P	[amol·s ⁻¹ ·x ⁻¹]	0.248 (0.020)				
FNS_P	[amol·s ⁻¹ ·x ⁻¹]	0.376 (0.030)				
$FNSGp_P$	[amol·s ⁻¹ ·x ⁻¹]	0.450 (0.036)				
$FNSGp_E$	[amol·s ⁻¹ ·x ⁻¹]	0.766 (0.066)				
Indirect ca	Indirect calorimetry					
RMR (kca	1289.4 (31.8)					
Cardiopulmonary exercise tests						
peak V ₀₂	[mL·kg ⁻¹ ·min ⁻¹]	24.9 (1.0)				
peak $V_{\rm CO2}$	[mL·kg-1·min-1]	27.4 (1.3)				
peak RER		1.10 (0.02)				
Body composition						
BMI		26.6 (0.7)				
body tota	29.2 (1.4)					
body tota	39.8 (9.4)					
body tota	40.6 (1.1)					
trunk-on	38.5 (1.3)					

Maximal Uncoupled Respiration, and Max ETS (Mahapatra 2018). We compared these measures of platelet respirometry to resting metabolic rate (RMR), as well as body composition measures, such as BMI, fat mass, and lean mass. We also examined exercise fitness tests including peak V_{02} , peak V_{C02} , and peak respiratory exchange ratio (RER).

Table 2. Correlations of platelet respirometry with RMR, fitness, and body comp.

	Platelet respiration						
	F_P	FN_P	FNS_P	$FNSGp_P$	$FNSGp_E$		
Calorimetry							
resting metabolic rate (RMR)	.409*	.537**	.455*	.436*	.472*		
Cardiopulmonary exercise tests							
peak V_{02}	.076	.146	.208	.243	.249		
peak V _{CO2}	.205	.268	.325	.351	.368		
peak RER	.445*	.475*	.483*	.480*	.517**		
Body composition							
BMI	147	069	170	223	175		
body total fat mass	057	005	113	167	110		
body total lean mass	.097	.272	.130	.078	.178		
body total fat percent	171	200	278	310	298		
trunk-only fat percent	155	185	242	274	239		

Pearson coefficients of correlation *r* values. * $p \le 0.05$; ** $p \le 0.01$.

adjusted BMI

Peak RER

adjusted body total fat percent



.546**

.488*

.496*

Table 3. Adjusted correlations of platelet respirometry with RMR and peak RER								
		Platelet respiration						
	F_P	FN_P	FNS_P	$FNSGp_P$	$FNSGp_E$			
RMR								
unadjusted	.409*	.537**	.455*	.436*	.472*			
adjusted age	.426*	.541**	.452*	.429*	.456*			

.599**

.556**

.554**

.482*

.554**

.464*

.475* unadjusted .445* .483* .480* .517** adjusted age .473* .478* .470* .460* .481* .470* .478* adjusted BMI .443* .475* .509** adjusted body total fat percent .432* .455* .463* .459*

.513**

.431*

Pearson coefficients of correlation *r* values. * $p \le 0.05$; ** $p \le 0.01$

3.2. Relationships between platelet bioenergetics and resting metabolic rate

We identified positive correlations between RMR and five measures of platelet respiration (Table 2; Figure 2). Controlling for age, BMI, and percent body fat, indicate that these covariates had little influence on the relationship between RMR and platelet respiration (Table 3).

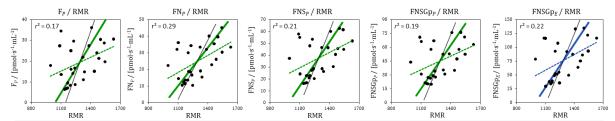


Figure 2. Correlations of platelet bioenergetics and RMR. Y/X regression lines (dashed; lowest slope using ordinary least squares), ordinate projection of X/Yabscissal regression lines (dotted), and mean regression lines (full). Coefficients of determination r^2 are independent of axis inversion. See Gnaiger (2021).

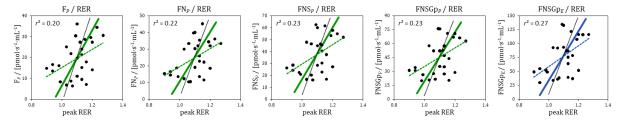


Figure 3. Correlations of platelet bioenergetics and peak RER. Regression lines are calculated as in Figure 2.

3.3. Relationships between platelet bioenergetics and measures of exercise fitness

We also identified positive correlations between peak RER and five measures of platelet respiration (Table 2; Figure 3) using Pearson correlation analyses. Peak V_{02} and peak $V_{\rm CO2}$ were not correlated with measures of platelet respiration. Controlling for age, BMI, and percent body fat, indicate that these covariates had little influence on the relationship between peak RER and platelet respiration (Table 3).



3.4. Relationships between platelet bioenergetics and measures of body composition

Relationships between fat % and FNS_P (r=-0.278), maximum OXPHOS capacity (FNSGp_P; r=-0.31), and maximum ET capacity (FNSGp_E; r=-0.298) are reported in Table 2. Similar relationships were found between trunk fat % and FNS_P (r=-0.242), FNSGp_P (r=-0.274) and FNSGp_E (r=-0.239). While these correlations are trending, all have p-values greater than 0.05.

4. Discussion

This study examines platelet mitochondrial function in healthy women over 60 years of age. We found that peak RER and RMR are both positively correlated to all measures of platelet bioenergetic function examined, independent of body composition.

Age-related mitochondrial decline and its relationship to physical function has predominantly been studied in skeletal muscle due to the strong association between sarcopenia and physical function decline. Here, we build on prior literature by focusing on blood-based bioenergetics and the ability of blood cells to report on physical health. Our data are in line with reports that skeletal muscle bioenergetics are associated with RMR (Choi et al 2016; Tyrrell et al 2015). We also found a strong relationship between platelet bioenergetics and peak RER, which has not been previously reported (Knuiman et al 2021). Together, these new findings contribute to our understanding of how blood-based bioenergetic profiling relates to physical fitness and exercise physiology.

RER is the ratio of carbon dioxide output to oxygen uptake ($V_{\rm CO_2}/V_{\rm O_2}$) and is typically measured by gas exchange at the mouth. At higher exercise intensity, increased lactate buildup associated with anaerobic metabolism contributes to a disproportionate increase in $V_{\rm CO_2}$ that brings RER to values >1 (Balady et al 2010; Milani 2006). Thus, peak RER can be used as a reliable, quantitative measure of maximal exercise effort. Our findings indicate that platelet maximum ET capacity (FNSGp_E) most strongly correlates to maximal exercise effort. Additional measures of platelet respirometry, such as fatty acid oxidation as well as individual complex function, also correlate positively to maximal exercise effort.

Interestingly, we did not observe an association between platelet bioenergetics and peak V_{02} , which has been previously found to be related to skeletal muscle bioenergetics (Knuiman et al 2021; Coen et al 2013; Distefano et al 2017; Gonzalez-Freire 2018). While this finding has been previously reported in skeletal muscle, but not blood, it should be noted that the women enrolled in this study exhibited relatively similar levels of fitness to each other. This small dynamic range and small sample size suggest that we were not adequately powered to observe this potential relationship. Future studies should be designed to determine if platelet bioenergetics are associated with RMR and exercise capacity in both men and women, in addition to over a larger age range to analyze how these relationships may change over lifespan and healthspan.

Overall, these findings suggest that energy expenditure during rest and physical activity are related to systemic mitochondrial function. These data can be used as a foundation to study how potential interventions, such as diet and exercise, may lead to improvements in mitochondrial function, resting metabolic rate, and other measures of physical fitness. Blood-based bioenergetic profiling, a minimally-invasive technique, can be used to track improvements and changes in exercise fitness in clinical studies.



5. Conclusions

In this study, we report positive correlations across five measures of platelet respiration with both peak RER and RMR, thus contributing to a growing body of evidence indicating that this minimally-invasive evaluation of mitochondrial function relates to physical health. The ability of blood cells to recapitulate skeletal muscle bioenergetics and predict exercise fitness suggests that systemic bioenergetic capacity may play a key role in physical fitness. Blood-based bioenergetic profiling may serve as a reliable biomarker of mitochondrial health among older adults and may be utilized to test efficacy and identify targets of interventions designed to promote the health and well-being of older adults.

Acknowledgements

We thank Oroboros Instruments for technical support and the loan of O2k respirometers for the conduct of this study. This project was supported by grants awarded to Dr. Molina from the National Insitutes for Aging (R01 AG054523, R21 AG051077) and the American Heart Association (15MCPRP25680019).

References

- Albouaini K, Egred M, Alahmar A, Wright DJ (2007) Cardiopulmonary exercise testing and its application. Postgrad Med J 83:675-82. doi:10.1136/hrt.2007.121558
- Balady GJ, Arena R, Sietsema K, et al (2010) Clinician's guide to cardiopulmonary exercise testing in adults. Review-article. Circulation 122:191-225. doi:10.1161/CIR.0b013e3181e52e69
- Braganza A, Corey CG, Santanasto AJ, Distefano G, Coen PM, Glynn NW, Nouraie SM, Goodpaster BH, Newman AB, Shiva S (2019) Platelet bioenergetics correlate with muscle energetics and are altered in older adults. JCI Insight 5. doi: 10.1172/jci.insight.128248
- Choi S, Reiter DA, Shardell M, Simonsick EM, Studenski S, Spencer RG, Fishbein KW, Ferrucci L (2016) ³¹P magnetic resonance spectroscopy assessment of muscle bioenergetics as a predictor of gait speed in the Baltimore Longitudinal Study of Aging. J Gerontol A Biol Sci Med Sci 71:1638-45. doi: 10.1093/gerona/glw059
- Coen PM, Jubrias SA, Distefano G, Amati F, Mackey DC, Glynn NW, Manini TM, Wohlgemuth SE, Leeuwenburgh C, Cummings SR, Newman AB, Ferrucci L, Toledo FG, Shankland E, Conley KE, Goodpaster BH (2013) Skeletal muscle mitochondrial energetics are associated with maximal aerobic capacity and walking speed in older adults. J Gerontol A Biol Sci Med Sci 68:447-55. doi: 10.1093/gerona/gls196
- Distefano G, Standley RA, Dubé JJ, et al (2017) Chronological age does not influence ex-vivo mitochondrial respiration and quality control in skeletal muscle. J Gerontol A Biol Sci Med Sci 72:535-42. doi:10.1093/gerona/glw102
- Edwards LM, Kemp GJ, Dwyer RM, Walls JT, Fuller H, Smith SR, Earnest CP (2013) Integrating muscle cell biochemistry and whole-body physiology in humans: ³¹P-MRS data from the InSight trial. Sci Rep 2013:1182. doi: 10.1038/srep01182
- Gnaiger E (2020) Mitochondrial pathways and respiratory control. An introduction to OXPHOS analysis. 5th ed. Bioenerg Commun 2020.2. https://doi.org/10.26124/bec:2020-0002
- Gnaiger E (2021) Bioenergetic cluster analysis mitochondrial respiratory control in human fibroblasts. MitoFit Preprints 2021.8. https://doi.org/10.26124/mitofit:2021-0008
- Gonzalez-Freire M, Scalzo P, D'Agostino J, Moore ZA, Diaz-Ruiz A, Fabbri E, Zane A, Chen B, Becker KG, Lehrmann E, Zukley L, Chia CW, Tanaka T, Coen PM, Bernier M, de Cabo R, Ferrucci L (2018) Skeletal muscle ex vivo mitochondrial respiration parallels decline in vivo oxidative capacity, cardiorespiratory fitness, and muscle strength: The Baltimore Longitudinal Study of Aging. Aging Cell 17. doi: 10.1111/acel.12725
- Hägg S, Jylhävä J (2021) Sex differences in biological aging with a focus on human studies. eLife 10:e63425. https://doi.org/10.7554/eLife.63425
- Knuiman P, Straw S, Gierula J, et al (2021) Quantifying the relationship and contribution of mitochondrial



- respiration to systemic exercise limitation in heart failure. ESC Heart Fail 8:898-907. doi:10.1002/ehf2.13272
- Larsen FJ, Schiffer TA, Sahlin K, Ekblom B, Weitzberg E, Lundberg JO (2011) Mitochondrial oxygen affinity predicts basal metabolic rate in humans. FASEB J 25:2843-52. doi:10.1096/fj.11-182139
- Lenaz G, D'Aurelio M, Merlo Pich M, et al (2000) Mitochondrial bioenergetics in aging. Biochim Biophys Acta 1459:397-404. doi:10.1016/s0005-2728(00)00177-8
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G (2013) The hallmarks of aging. Cell 153:1194-217. doi:10.1016/j.cell.2013.05.039
- Mahapatra G, Smith SC, Hughes TM, Wagner B, Maldjian JA, Freedman BI, Molina AJA (2018) Blood-based bioenergetic profiling is related to differences in brain morphology in African Americans with Type 2 diabetes. Clin Sci (Lond) 132:2509-18. doi: 10.1042/CS20180690
- Marzetti E, Calvani R, Cesari M, Buford TW, Lorenzi M, Behnke BJ, Leeuwenburgh C (2013) Mitochondrial dysfunction and sarcopenia of aging: from signaling pathways to clinical trials. Int J Biochem Cell Biol 45:2288-301. doi: 10.1016/j.biocel.2013.06.024
- McMurray RG, Soares J, Caspersen CJ, McCurdy T (2014) Examining variations of resting metabolic rate of adults: a public health perspective. Med Sci Sports Exerc 46:1352-8. doi:10.1249/MSS.000000000000232
- Milani RV, Lavie CJ, Mehra MR, Ventura HO (2006) Understanding the basics of cardiopulmonary exercise testing. Mayo Clin Proc 12:1603-11. doi: 10.4065/81.12.1603
- Nicklas BJ, Brinkley TE, Houston DK, Lyles MF, Hugenschmidt CE, Beavers KM, Leng X (2019) Effects of caloric restriction on cardiorespiratory fitness, fatigue, and disability responses to aerobic exercise in older adults with obesity: a randomized controlled trial. J Gerontol A Biol Sci Med Sci 74:1084-90. doi: 10.1093/gerona/gly159
- Nguyen QL, Wang Y, Helbling N, Simon MA, Shiva S (2019) Alterations in platelet bioenergetics in Group 2 PH-HFpEF patients. PLoS One doi:10.1371/journal.pone.0220490
- Ravussin E, Bogardus C (1989) Relationship of genetics, age, and physical fitness to daily energy expenditure and fuel utilization. Am J Clin Nutr 49:968–75. https://doi.org/10.1093/ajcn/49.5.968
- Short KR, Bigelow ML, Kahl J, Singh R, Coenen-Schimke J, Raghavakaimal S, Nair KS (2005) Decline in skeletal muscle mitochondrial function with aging in humans. Proc Natl Acad Sci U S A 102:5618-23 doi:10.1073/pnas.0501559102
- Sjövall F, Morota S, Asander Frostner E, Hansson Magnus J, Elmer E (2014) Cytokine and nitric oxide levels in patients with sepsis temporal evolvement and relation to platelet mitochondrial respiratory function. PLoS One 9:e97673.
- Tyrrell DJ, Bharadwaj MS, Van Horn CG, Kritchevsky SB, Nicklas BJ, Molina AJ (2015) Respirometric profiling of muscle mitochondria and blood cells are associated with differences in gait speed among community-dwelling older adults. J Gerontol A Biol Sci Med Sci 70:1394-9. doi: 10.1093/gerona/glu096
- Tyrrell DJ, Bharadwaj MS, Jorgensen MJ, Register TC, Molina AJ (2016) Blood cell respirometry is associated with skeletal and cardiac muscle bioenergetics: implications for a minimally invasive biomarker of mitochondrial health. Redox Biol 10:65-77. doi:10.1016/j.redox.2016.09.009
- Wang Z, Ying Z, Bosy-Westphal A, et al (2010) Specific metabolic rates of major organs and tissues across adulthood: evaluation by mechanistic model of resting energy expenditure. Am J Clin Nutr 92:1369-77. doi:10.3945/ajcn.2010.29885
- Zampino M, Semba RD, Adelnia F, Spencer RG, Fishbein KW, Schrack JA, Simonsick EM, Ferrucci L (2020) Greater skeletal muscle oxidative capacity is associated with higher resting metabolic rate: results from the Baltimore Longitudinal Study of Aging. J Gerontol A Biol Sci Med Sci 75:2262-8. doi: 10.1093/gerona/glaa071
- **Copyright:** © 2022 The authors. This is an Open Access preprint (not peer-reviewed) distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original authors and source are credited. © remains with the authors, who have granted MitoFit Preprints an Open Access publication license in perpetuity.

